



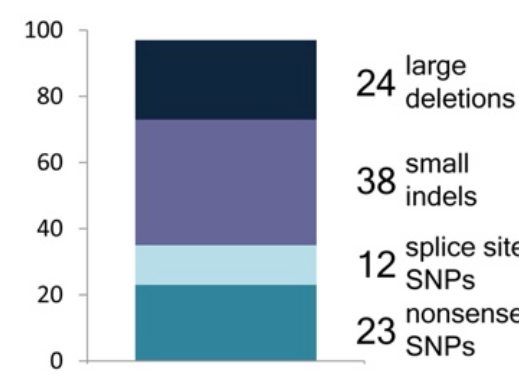
Mutations in DNA Sequencing Studies

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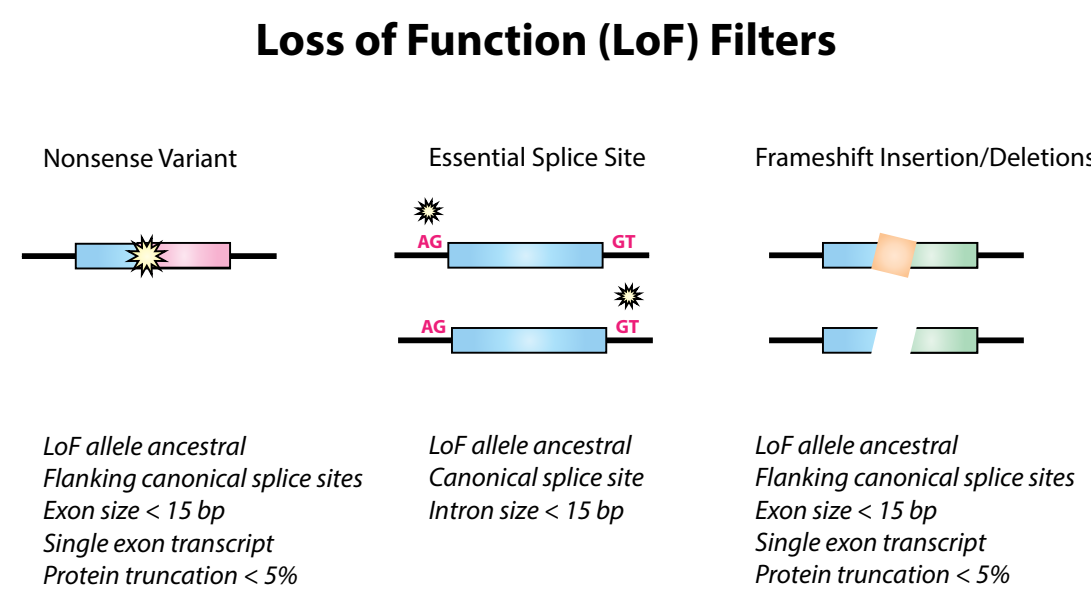
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Loss-of-Function Mutations in DNA Sequencing Studies

- Protein truncating variants (frameshift, stop gain, splice site) are commonly referred to as Loss-of-Function (LoF).
- A typical genome has ~100 annotated germline LoF variants.
- Annotated LoF variants may have a strong impact on disease outcome^{3,4,5}.



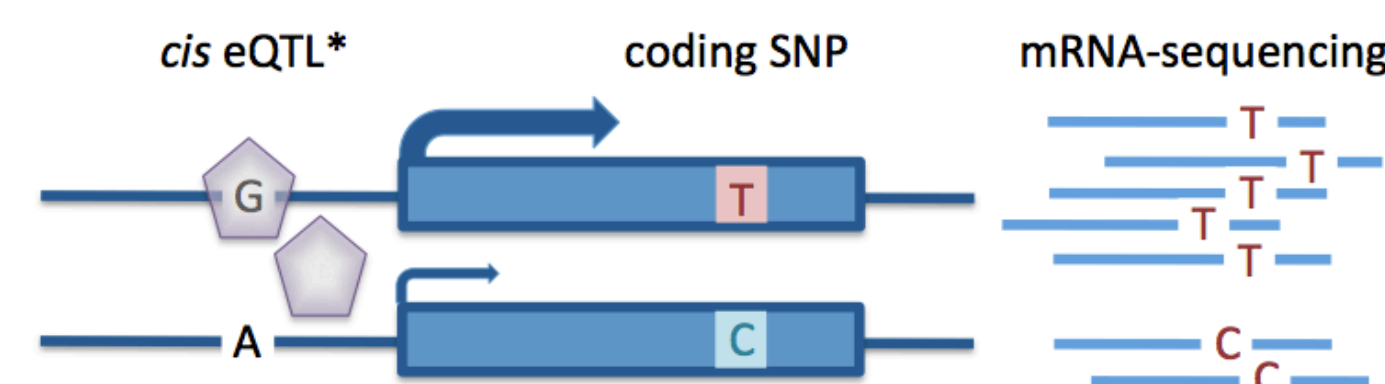
Annotation



- DNA Sequence variants annotated with ALOFT and PLINK/SEQ⁶ using Gencode version 12 transcripts.

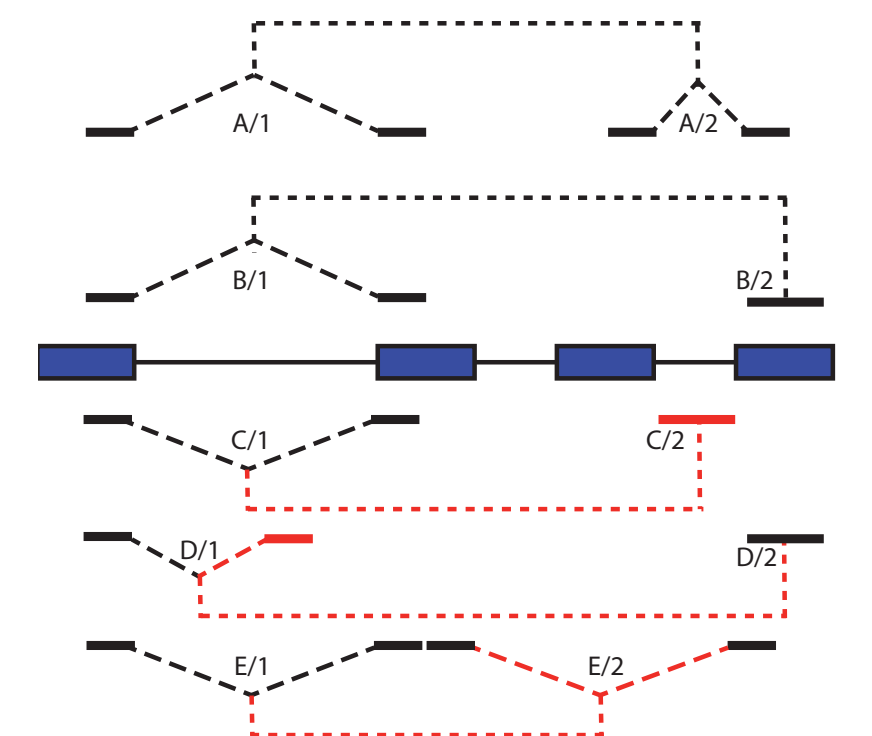
Methods

Allele Specific Expression (ASE)



- Read counts over all heterozygous sites of an individual partitioned by the allele.
- Binomial test to detect deviation from the expected ~50/50 ratio.

Splice Junction Quantifications



Two examples for valid, annotation-paired split-mappings (A and B) as well as three split-mappings that are incompatible with the annotation (C, D, E).

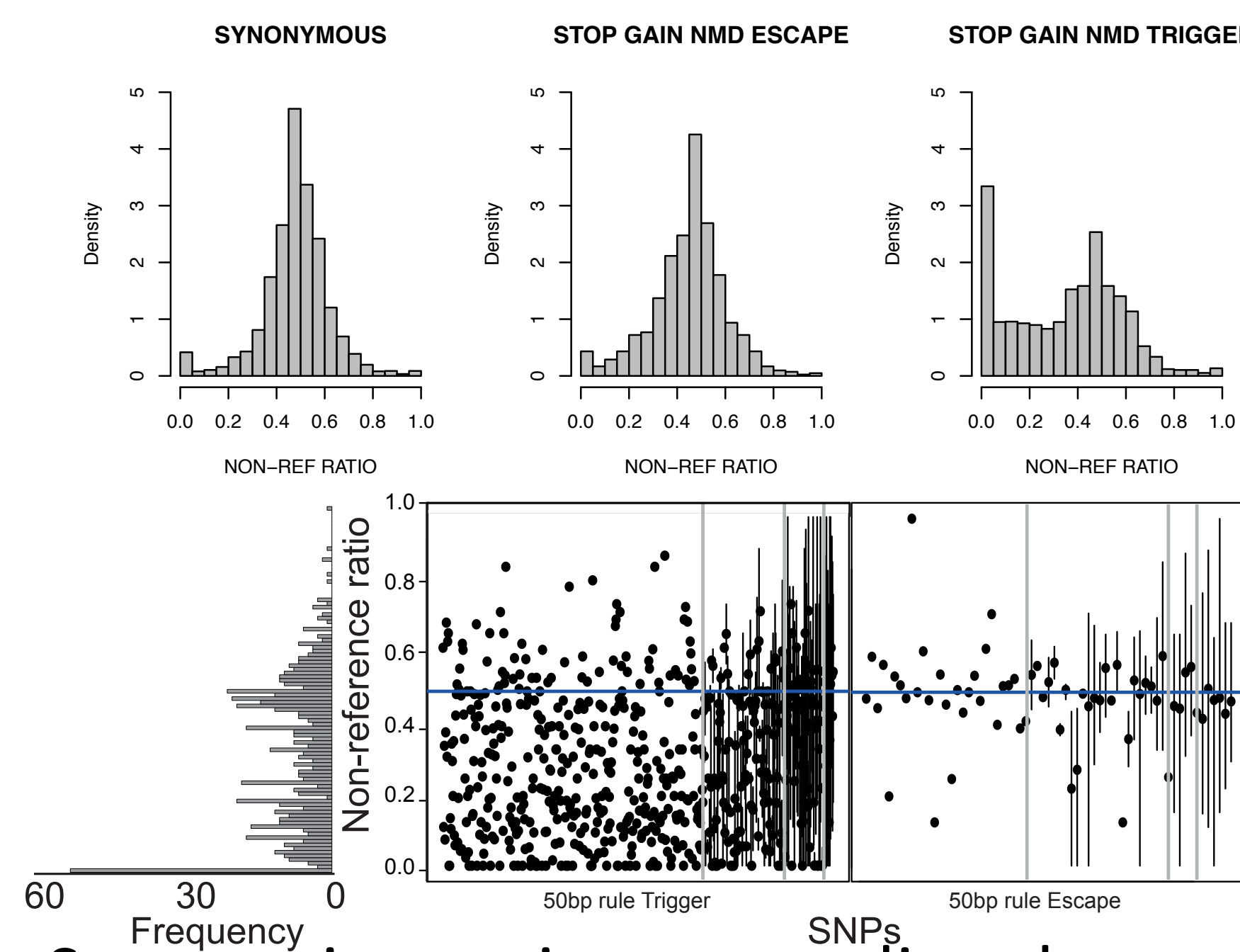
- Splice junction quantifications using annotation-compatible split-mappings.

Aims

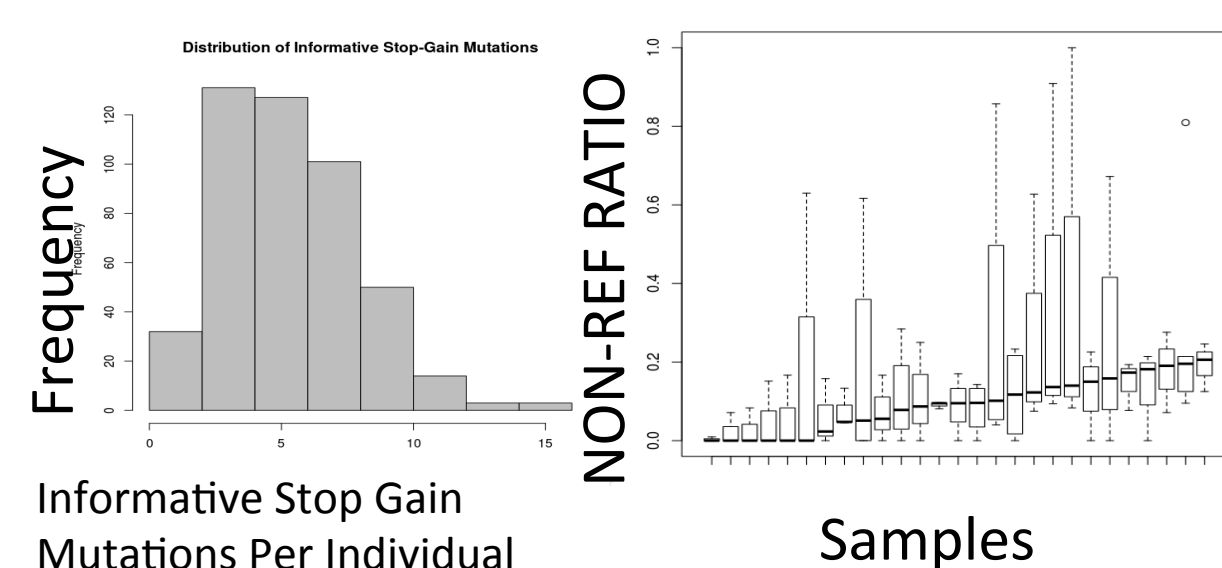
- Using RNASeq data to develop more precise description of functional effect for loss-of-function variants, e.g with respect to Nonsense Mediated Decay (NMD) and splicing.

Results

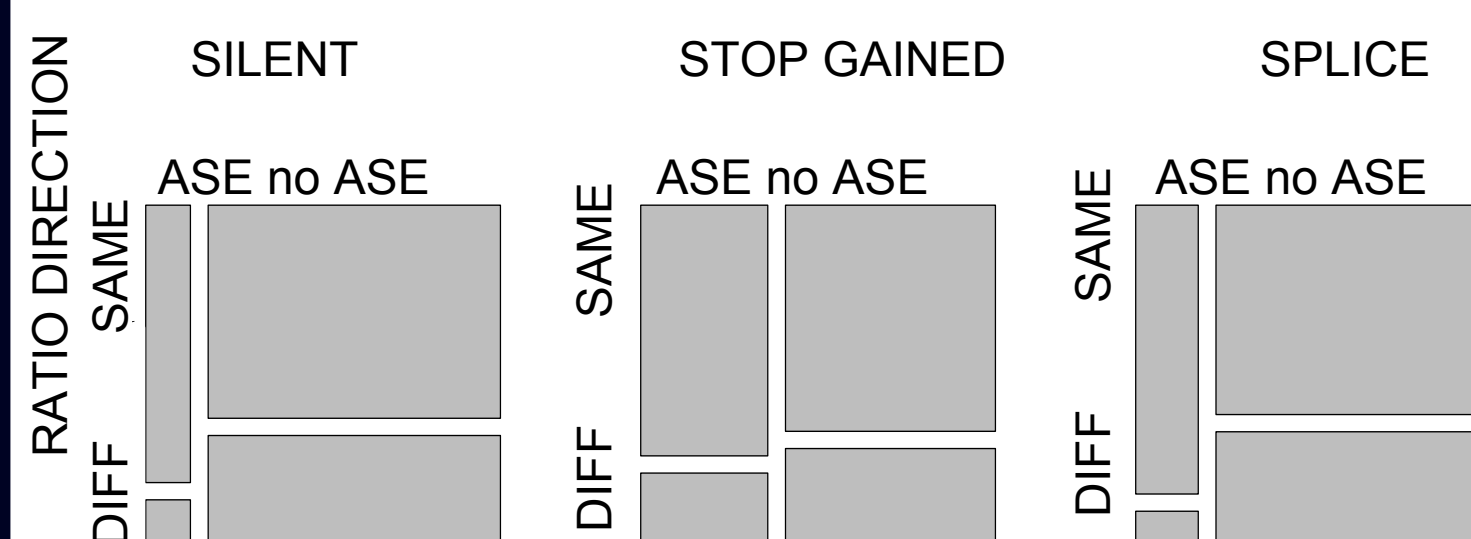
Stop Gain variants and ASE Signal



- Stop gain variants predicted to trigger nonsense-mediated decay (NMD) exhibit different ASE distribution from Synonymous variants and STOP gain variants predicted to escape NMD ($P < 2.2e-16$). Better predictions needed for NMD.

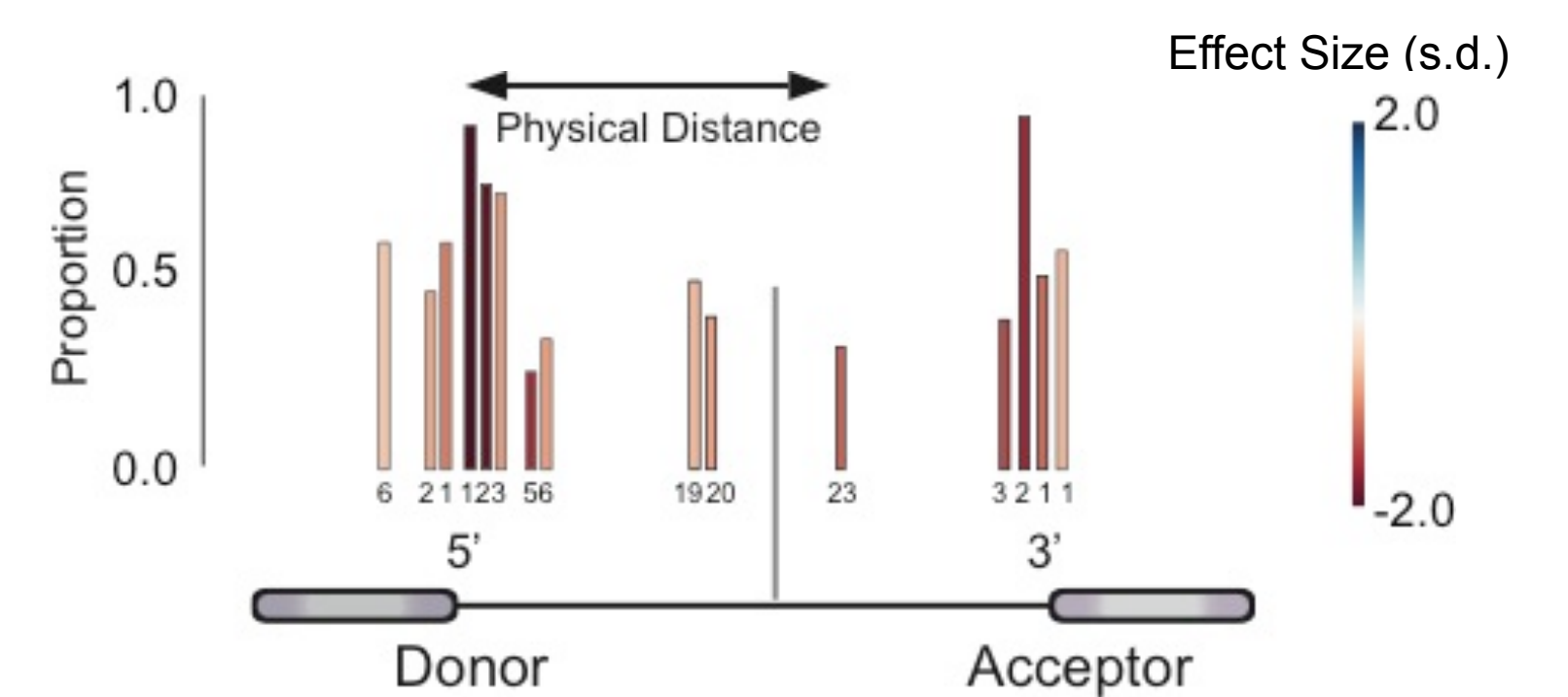


Substantial variance in ASE signal between individuals.

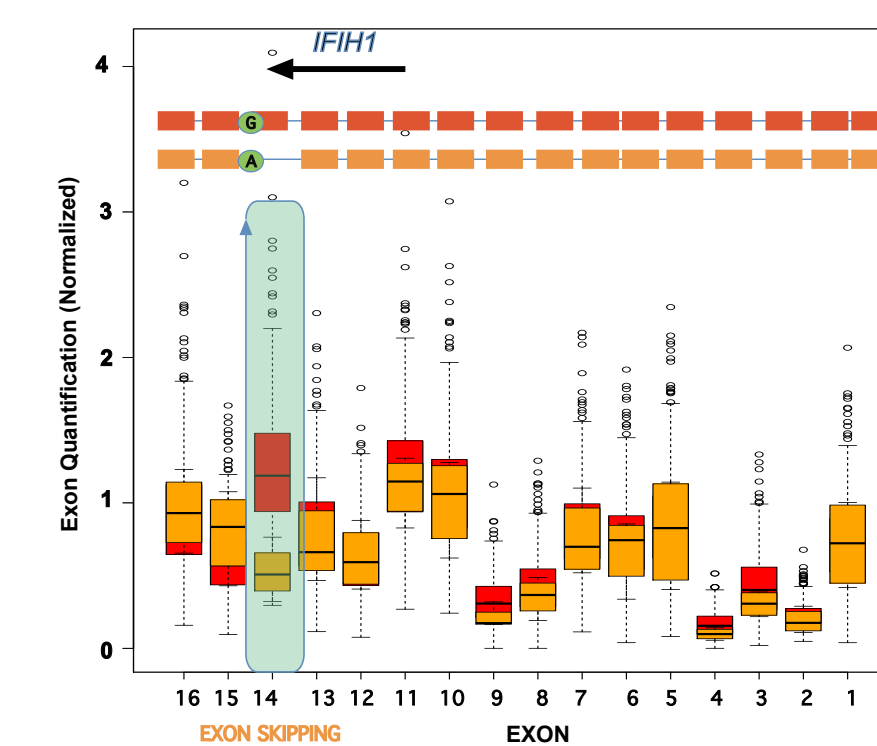


- ASE signals are not very consistent within a gene, but for stop gain variants the consistency is higher – i.e. they tend to affect the entire gene, more often than regulatory variation that only affect a subsection of the gene.

Variants near splice junctions



MCMC algorithm applied to splice junction quantifications highlight shared impact of rare variants near splice junctions.



- Rare protective splice variant c.IVS14+1G>A at *IFIH1* disrupts splicing of exon 14 and leads to exon skipping while escaping nonsense mediated decay.

Functional interpretation of LoF variants associated to disease.

References

¹MacArthur, D.G. et al. A Systematic Survey of Loss-of-Function Variants in Human Protein-Coding Genes. *Science* 2012.
²The 1000 Genomes Project Consortium. An integrated map of genetic variation from 1,092 human genomes. *Nature* 2012.
³Nejentsev, S. et al. Rare variants of *IFIH1*, a Gene Implicated in Antiviral Responses, Protect Against Type 1 Diabetes. *Science* 2009.
⁴Rivas, M.A. et al. Deep resequencing of GWAS loci identifies independent rare variants associated with inflammatory bowel disease. *Nature Genetics* 2011.
⁵Neale, B.M. et al., Sanders et al., O’Roak et al., 2012. *Nature* 2012.
⁶<http://atgu.mgh.harvard.edu/plinkseq/>; PLINK/SEQ v0.09. ALOFT MacArthur, D.G. et al.

mRNA Sequencing of 465 Samples from the 1000 Genomes Project

- RNA Sequencing done in seven institutes with Illumina TruSeq Protocol.
- Lymphoblastoid cell lines (LCL) from 465 samples.
- Genotypes from 1000 Genomes 27M total variants. 90% of samples in Phase 1, remaining imputed from Omni 2.5
- 660 informative stop gain variants; ~5000 informative variants near splice junctions.

	1000G Phase1 + Phase2	RNAseq
TSI	92 + 1	93
GBR	86 + 10	96
FIN	89 + 6	95
CEU	79 + 13	92
YRI	77 + 12	89
TOTAL	423 + 42	465

See Poster 567F for more information about the Geuvadis RNA Sequencing Project.